## Tissues, Pathology, and Diagnostic Microscopy

## LS.2.P045 Chemical-induced hemopoietic cell death: the protective action of melatonin

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Melatonin is a methoxyindole synthesized by the pineal gland. The hormone is secreted during the night and appears to play multiple roles in the human organism.

Melatonin is a powerful anti-oxidant in tissues [1] as well as within cells [2], with a fundamental role in ameliorating homeostasis in a number of specific pathologies. It acts both as a direct radical scavenger and by stimulating production/activity of intracellular anti-oxidant enzymes, i.e. superoxide dismutase and glutathione peroxidase as well as up-regulating their expression. Its function consists in the reduction of oxidative stress, i.e. molecular damage produced by reactive oxygen and nitrogen species [3].

A survey of literature shows that in leukocytes melatonin mainly exerts an anti-apoptotic role, as also demonstrated in an our previous work exposing hemopoietic cells to UVB rays [4].

In this work, some chemical triggers (hydrogen peroxide, etoposide and staurosporine), with different mechanism of action and able to increase radical oxygen species levels [5], have been chosen to induce cell death in U937 hemopoietic cell line. Cells were pre-treated with different melatonin concentrations (1 or 0.1 mM) and then exposed to chemical agents.

Potential melatonin anti-oxidant and anti-apoptotic effects were evaluated through morphological and statistical analyses, the latter by means of Tali Image-Based Cytometer, able to monitor cell viability as well as apoptosis presence. Samples, observed at scanning (SEM) and transmission electron microscopy (TEM), revealed a preserved morphology in control (Figure 1A-B) and melatonin alone (Figure 1C-D) conditions. After all chemical treatments typical apoptotic features appeared. Cells showed membrane blebs, nuclear cup-shaped masses, micronuclei, altered and swollen mitochondria, cytoplasmic vacuolisation and secondary necrosis (Figure 1E, F, I, L, O, P). A cell death decrease was observed in all experimental conditions pre-treated with melatonin (Figure 1G, H, M, N, Q).

In addition, in cells pre-incubated with melatonin (in both concentrations) and then exposed to chemicals, autophagic vacuoles appeared at TEM (Figure 1R).

Melatonin anti-apoptotic activity was also demonstrated by quantitative analyses. Supravital propidium iodide [6], used to evaluate cell viability, evidenced that melatonin administration significantly prevents cell death (Table 1). In particular, melatonin 100  $\mu$ M showed an elevated protection against apoptosis induced by hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), staurosporine and etoposide. Melatonin 1mM has moderate anti-oxidant and anti-apoptotic effects *vs* chemical treatments.

In conclusion, we found that in U937 cell line melatonin can be considered a powerful molecule in preventing chemical apoptotic cell death.

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**Figure 1.** U937 cells at SEM (A, C, E, G, I, M) and TEM (B, D, F, H, L, N,O,P,Q,R). A preserved morphology is evident in control (A,B) and melatonin (0.1mM) alone conditions (C,D). Cells treated with  $H_2O_2$  (E,F) etoposide (I,L) and staurosporine (O,P) reveal typical apoptotic patterns. Melatonin added to  $H_2O_2$  (G,H) etoposide (M,N) and staurosporine (Q) treated cells prevents cell death. In R, autophagic vacuoles appeared in melatonin added to etoposide. Bars: 2µm in A-D, O,Q; 5 µm in E-N; 1 µm in P; 0.5 µm in R.



**Table 1.** Cell viability percentage. Data have been are verified by t-test and appeared statistically significant (p < 0.05).